

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicant: Ebrahim ZANDI, et al.

Title: COMPOSITION AND METHOD
FOR RECONSTITUTING I κ B
KINASE IN YEAST AND
METHODS OF USING SAME

Appl. No.: 10/079,949

Filing Date: 2/19/2002

Examiner: Prouty, Rebecca E.

Art Unit: 1652

Confirmation Number: 6542

SUPPLEMENTAL DECLARATION UNDER 37 CFR SECTION 1.131

Commissioner for Patents
P.O. Box 1450
Alexandria, VA 22313-1450

Sir:

1. We, Ebrahim Zandi and Beth Schomer Miller, hereby declare as follows.
2. We are the Ebrahim Zandi and Beth Schomer Miller, who are named as co-inventors of the above-identified application.
3. That we conceived and reduced to practice in the United States the transformation of an IKK subunit gamma (γ) gene, an IKK subunit alpha (α) gene and/or an IKK subunit beta (β) gene into yeast and the separation from that yeast a substantially homogenous and biologically functional IKK protein complex prior to November 15, 2000, the online publication date of the literature article Li et al. (2001) "Role of IKK γ /NEMO in Assembly of the I κ B Kinase Complex"

Journal of Biological Chemistry 276(6):4494-4500. Attached hereto is Exhibit A, a copy of pages from laboratory notebooks recorded by Beth Schomer Miller working under our direct control and supervision showing a reduction to practice wherein the activity of a purified IKK complex from yeast transformed with either IKK β , IKK $\beta\gamma$, or IKK $\alpha\beta\gamma$ compared to mammalian IKK complex isolated from control Hela cells or TNF stimulated HELA cells was determined. These experimental results demonstrate that a yeast cell was transformed with an IKK subunit gamma (γ) gene, an IKK subunit alpha (α) gene and/or an IKK subunit beta (β) gene. The yeast was then grown and a substantially homogenous and biologically functional IKK protein complex was separated from the yeast.

4. That the documents in Exhibit A, which relates to the aforementioned actual reduction to practice, are exact and true copies. All personal information, including names and dates have been redacted from the documents, but all dates are prior to November 15, 2000.

5. Attached hereto is Exhibit B. Exhibit B is a typed version of Exhibit A and a true and exact representation of the handwritten information of Exhibit A.

6. As specifically identified on pages 1 and 9 of Exhibit B, the purpose of the experiment was to compare the activity of recombinantly produced IKK complexes that were isolated from yeast transformed with IKK subunits alpha, beta, and gamma ($\gamma\alpha\beta\gamma$), subunits beta and gamma ($\gamma\beta\gamma$), subunit beta ($\gamma\beta$), to IKK complexes isolated from non-stimulated Hela cells (HNS) and TNF-stimulated Hela cells (TNF). The remaining pages set forth the experimental protocols, the resulting activity of the isolated IKK complexes and the amount of IKK β subunit present in the isolated IKK complexes. The gels shown on pages 4, 5, 10 and 11 of Exhibit B demonstrate that the activity of recombinantly produced IKK complex isolated from yeast transformed with IKK subunits alpha, beta, and gamma ($\gamma\alpha\beta\gamma$) is higher than purified IKK complex from non-stimulated HeLa cells and the same or slightly higher than purified activated IKK complex from TNF-stimulated Hela cells. Similar gels and results are show in Figure 3 and on page 15, lines 21 to 27 of the above-identified application.

Atty. Dkt. No. 064189-0501

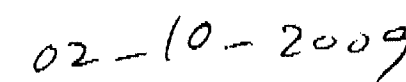
7. We further declare that all statements made herein of our own knowledge are true and that all statements made on information and belief are true; and further that all statements made herein are made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such false statements may jeopardize the validity of legal decisions of any nature based on them.

Respectfully submitted,

Ebrahim Zandi



Signature:



Date:

Beth Schomer Miller

Signature:

Date:

Atty. Dkt. No. 064189-0501

7. We further declare that all statements made herein of our own knowledge are true and that all statements made on information and belief are true; and further that all statements made herein are made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such false statements may jeopardize the validity of legal decisions of any nature based on them.

Respectfully submitted,

Ebrahim Zandi

Signature:

Date:

Beth Schomer Miller

Beth Schomer Miller

Signature:

10 Feb 09

Date:

Exhibit A

Purpose: to compare IRL activity in
y LBS yB HNS TNF

BUT GF LBS 8-25 good HA signal in 2 sec exp.
SML Fr 10 or 11

BUT GF LBS good LBS 1 min SML Fr 10-11
1/2 y LBS HA

good signal 15 sec similar to

1/2 of y LBS Fr 10

BUT B HA detect same Fr 15 after 1 min (15)

HNS GF

HNS B detected in 10 + 11 aft 1 min 20
TNF weakly detected in 10 + 11 40 min 20

INP. identical.

Hugo's westerns were also poor for detection of
B in B alone + LBS in his assay.

I'll have to play around with amounts

HNS Q20 + TNF Q20 were separated by gel filtration
INP. could detect S-15 by LBS + y western
in 15 sec.

Less present than S1 y LBS

Put fractions in gel filtration → ~10 fold dilution

would need to use 150 for same amt.

Concentrate 150(10) + 150(11)
300 → 30

use: 5, 10, 15

B-HA fraction IS G.F.
3' + 12' 1x KA
5' + 10' 1x
10' + 5' 1x

BX -HA Fr 10
3' + 12' 1x
5' + 10' 1x
10' + 5' 1x

2 BX Fr 10-11
3' + 12' 1x
5' + 10' 1x

HNS G 20 → Sp 6 GF 10-11 17
200 + 200 → 400
use 5, 10, 15

TNF G 20 → Sp 6 GF 10-11
200 + 200 → 400
use 5, 10, 15

Load 35% each

1 empty ✓
2 empty ✓
3 B3 ✓
4 S ✓
5 D ✓
6 B3 ✓
7 S ✓
8 D ✓
9 XBS 3 ✓
10 S ✓
11 HNS 5 ✓
12 D ✓
13 S ✓
14 TNF 5 ✓
15 D ✓
16 S ✓

all
correctly
loaded

UL-TAPPER MC
SD, KD

put
300 1x kinase
buffer in
bottom
↓
prevent drying
risk

Should recover

≤ 40% rehydrate
renew

20% 25% HNS
at 15' 1x

40% TNF

1. aliquot extract + buffer according to table
2. add 30% kinase cocktail. Inc 30' 30°C
3. add 7x 1x SDS PAGE, heat
4. Load 10l. gel

only 62 added

Cocktail - 15
10x Kinase 45 ✓
20mm DTT 45 ✓
200mm ATP 45 ✓
0.5mg/ml GPT-His 30 ✓
8 ATP 7.5 ✓
H2O 277.5 ✓

31 90-58
44 58

[REDACTED] Purpose: to compare activity of
yB vs yB⁺ vs yAB⁺ vs HNS vs TUF

10% gel (10-15)
30% acryl
8.8
H₂O
APS
TEMED

5.2
3.75
6.25
~~200~~ 100
~~210~~ 10

Stack
1.05
1.9 (6.8)
4.5
75
10

File/Range: D:\Users\1012bsm.gel / 0.000-45853 Counts / 1.000000

User Name: phospho

Image Name: D:\Users\1012bsm.gel

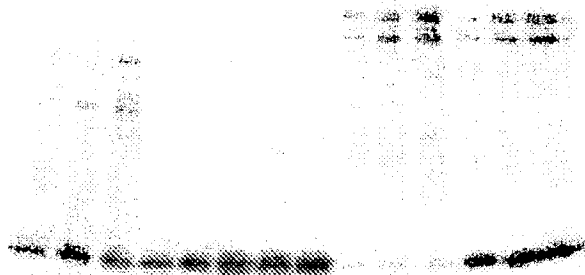
Image Comment: yeast b bg abg HNS TNF-Hela
scanned 9:13 am to 2:05 pm

Present Date/Time: [REDACTED]

Scan Date/Time: [REDACTED]

Prep. Date/Time: [REDACTED]

4/5			4/5/8			4/5/8			HNS			TNF		
3	5	10	3	5	10	3	5		3	5	10	3	5	10
									5	10	15	5	10	15



↑

↑ ↑
range 1-10,000

File/Range: D:\Users\1012bsm.gel / 0.000-45853 Counts / 1.000000

User Name: phospho

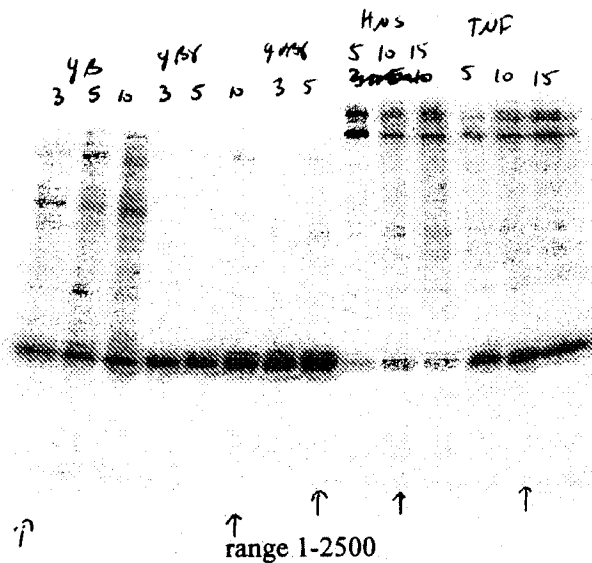
Image Name: D:\Users\1012bsm.gel

Image Comment: yeast b bg abg HNS TNF-Hela
scanned 9:13 am to 2:05 pm

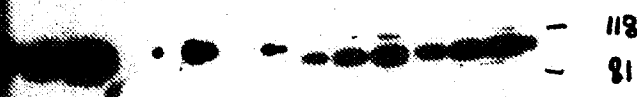
Present Date/Time: [REDACTED]

Scan Date/Time: [REDACTED]

Prep. Date/Time:



10 3 5 10 3 5 5 10 15 5 10 15
 207



118
 81
 52.5
 34
 30



207
 118
 81
 52.5
 34
 30



207
 118
 81
 52.5
 34
 30

113 4161 4161 1113 1113
 55 10 35 10 35 5 10 15 5 10 15

- 207
 - 118
 - 81
 - 52.5
 - 26
 - 10 5' up.



1113
 1113 1113 1113
 1113 1113
 1113 1113

1113 1113
 1113 1113

5 10 5 5 10 3 5 5 10 15 5 10 15 - 207

-110
-81

-525
W: 21145

1:500
-30

-30

6/12

Purpose: to compare IKK activity in
YB vs YBR vs YABR vs HNS vs TNF-Hen
repeat of 10-11 with attempt to use more similar amounts

HNS + TNF (Q20 → sup6 GF 10+11)

Put 300 μ 1x Kinase buffer in bottom to prevent drying.

Top: 200 μ sup6 GF 0 + 200 μ sup6 GF 11

Recover \sim 40 μ + adjust vol. to 40 μ
(1x KA)

B-HA fraction 15

Tube/lane

2 0.5 μ
3 1 μ
4 2 μ

5 B8 - \checkmark
6 \checkmark
7 \checkmark 21 μ

8 YBR \checkmark
9 \checkmark
10 \checkmark 21 μ

11 HNS \checkmark
12 \checkmark
13 \checkmark 15 μ

14 TNF \checkmark
15 \checkmark
16 \checkmark 15 μ

17 mw

21
35
56

all loaded
correctly

40 μ
lane.

5 μ + 45 μ 1x
1:10 in 1x kinase
 \checkmark 5 μ + 10 μ 1x
 \checkmark 10 μ + 11 μ 1x
 \checkmark 20 μ + 1 μ 1x
 \checkmark 5 μ + 14 μ 1x
 \checkmark 10 μ + 7 μ 1x
 \checkmark 15 μ + 0 μ

1. Aliquot extract +
buffer

2. Add ~~35 μ~~ 35 μ Kinase
cocktail Inc 30' 30 $^{\circ}$ C

3. Add ~~10 μ~~ 10 μ 1x
SDS PAGE
Heat 95 $^{\circ}$ C 5'

4. Load 10 μ gel
(40 μ)

Cocktail	16 samples	+	4
10x Kinase	48 μ		12 μ
20mM DTT	48 μ		12 μ
200mM ATP	48 μ		12 μ
GST-146	32 μ		8 μ
γ -P ATP	8 μ	906-58	2 μ
H ₂ O	296 μ		74 μ
	480		120

10 DTT
20 mM DTT
.02m 1m + .98 L H₂O

File/Range: D:\Users\1017bsm.gel / 0.000-45853 Counts / 0.814331

User Name: phospho

Image Name: D:\Users\1017bsm.gel

Image Comment: 2 experiments

1. 3 M urea GF column fractions (concentrated)

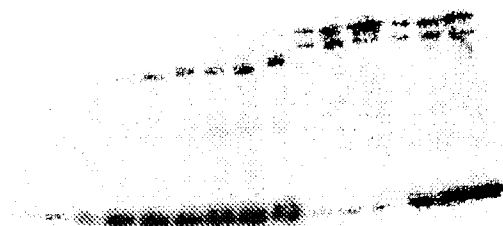
2. yeast b, bg, abg, HNS, TNF stim Hela

Present Date/Time:

Scan Date/Time:

Prep. Date/Time:

0.5 1 2 7 14 21 7 14 21 5 10 15 5 10 15
y b y b y b HNS TNF-Hela



scale 1-2500

y b

y b

HNS

TNF

11 yeast

File/Range: D:\Users\1017bsm.gel / 0.000-45853 Counts / 0.814331

User Name: phospho

Image Name: D:\Users\1017bsm.gel

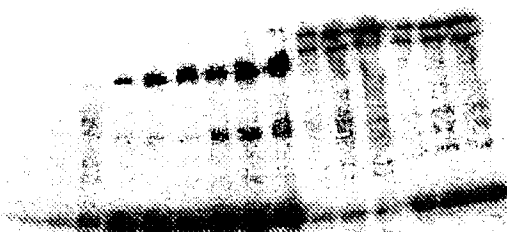
Image Comment: 2 experiments

1. 3 M urea GF column fractions (concentrated)
2. yeast b, bg, abg, HNS, TNF stim Hela

Present Date/Time:

Scan Date/Time:

Prep. Date/Time:



scale 1-250

90 401 4201 HNS TAF
 5-27-01 1-2 1-2 6-2-15 5-2-16



5/11/11
 KA,

1:50
 0W



1' up.

1. The first step in the process is to identify the problem. This involves gathering information about the situation and understanding the needs of the stakeholders involved.

—



5

2

6

10

e

—

2

—

1

1

10

Exhibit B

Purpose: to compare IKK activity in $\gamma\alpha\beta\gamma$ $\gamma\beta\gamma$ $\gamma\beta$ HNS TNF

Beth $\beta\gamma$ 8-25 good HA signal in 2 sec exp.
GF 5 μ L Fr 10 or 11

Beth $\alpha\beta\gamma$ good $\alpha+\gamma$ 1 min 5 μ L Fr 10-11
GF 1 λ $\gamma\alpha\beta\gamma$
HA good signal 15 sec similar to
1 λ of _____ $\gamma \rightarrow \beta\gamma$ Fr 10

Beth β HA detect some Fr 15 after 1 min (15 λ)

HNS GF

HNS β detected in 10 + 11 aft 1 min	20 λ
TNF weakly detected in 10 + 11 40 min	20 λ

INP identical

Hugo's westerns were also poor for detection of
 β in β alone + $\alpha\beta\gamma$ in his assays.
I'll have to play around with amounts

HNS Q20 + TNF Q20 were separated by gel filtration
Inp. could detect 5 – 15 λ by α IKK $\beta + \gamma$ Western in 15 sec.

Less present than 5 λ γ IKK

Put fractions in gel filtration \rightarrow ~ 10 fold dilution
would need to use 150 λ for same amt.

Concentrate 150(10) + 150(11) $\lambda \rightarrow$ 30 λ

use: 5 . 10 . 15

β -HA fraction 15 G.F.

3 λ + 12 λ 1x KA

5 λ + 10 λ 1x

10 λ + 5 λ 1x

$\beta\gamma$ -HA Fr 10

3 + 12 λ 1x

5 + 10 λ 1x

10 + 5 λ 1x

$\alpha\beta\gamma$ Fr 10-11

3 + 12 λ 1x

5 + 10 λ 1x

HNS Q 20 \rightarrow sup 6 GF 10 + 11

200 + 200 \rightarrow 40 λ

use 5, 10, 15 λ

+ +

10 λ 5 λ 1x

TNF Q 20 \rightarrow sup 6 GF 10 + 11

200 + 200 \rightarrow 40 λ

use 5, 10, 15 λ

+ +

10 λ 5 λ

1x 1x

Load 35 λ each

1. empty

2. empty

3. β 3

4. 5

5. 10

6. $\beta\gamma$ 3

7. 5 All correctly loaded

8. 10

9. $\alpha\beta\gamma$ 3

10. 5

11. HNS 5

12. 10

13. 15

14. TNF 5

15. 10

16. 15

17. mw ULTA FREE MC
30, KD

Put 300 λ 1x kinase
buffer in
bottom

\downarrow
prevent drying
filter

Should recover
 \leq 40 λ retentate
recover
25 λ HNS
add 15 λ 1x
40 λ TNF

1. Aliquot extract + buffer according to table

2. Add 30 λ Kinase cocktail Inc 30' 30°C

3. Add 9 λ 6x SDS PAGE, heat

4. Load 10% gel Only 6 λ added

Cocktail – 15

10x Kinase 45

20mm DTT 45

200 μ m ATP 45

0.5 mg/ μ l Gst - I κ B α 30 λ

γ ATP 7.5 λ 906-58 3 λ 906-57

H₂O 277.5 4.5 λ 58

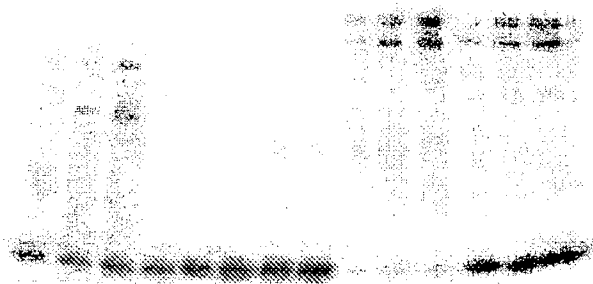
Purpose: to compare activity of
 $\gamma\beta$ vs $\gamma\beta\gamma$ vs $\gamma\alpha\beta\gamma$ vs HNS vs TNF

10% gel (10-10)		Stack
30% acryl	5ml	1.05
8.8	3.75	1.9 (6.8)
H ₂ O	6.25	4.5
APS	100	75
TEMED	10	10

File/Range: D:\Users\1012bsm.gel / 0.000-45853 Counts / 1.000000
 User Name: phospho
 Image Name: D:\Users\1012bsm.gel
 Image Comment: yeast b bg abg HNS TNF-Hela
 scanned 9:13 am to 2:05 pm

Present Date/Time: XXXXXXXXXX
 Scan Date/Time: XXXXXXXXXX
 Prep. Date/Time: XXXXXXXXXX

yβ			yβγ			yαβγ			HNS			TNF		
3	5	10	3	5	10	3	5	10	5	10	15	5	10	15

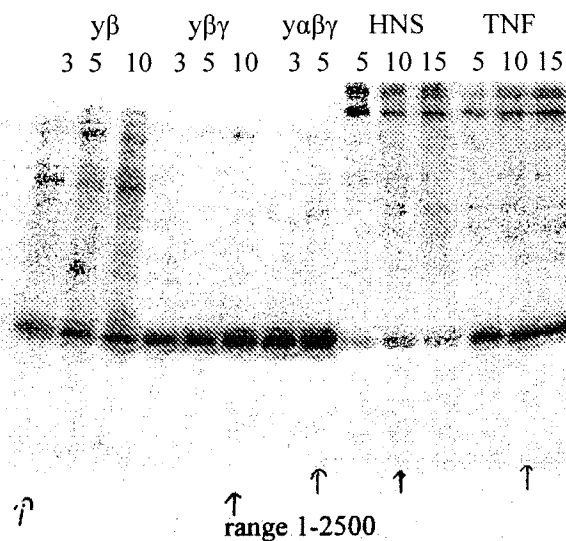


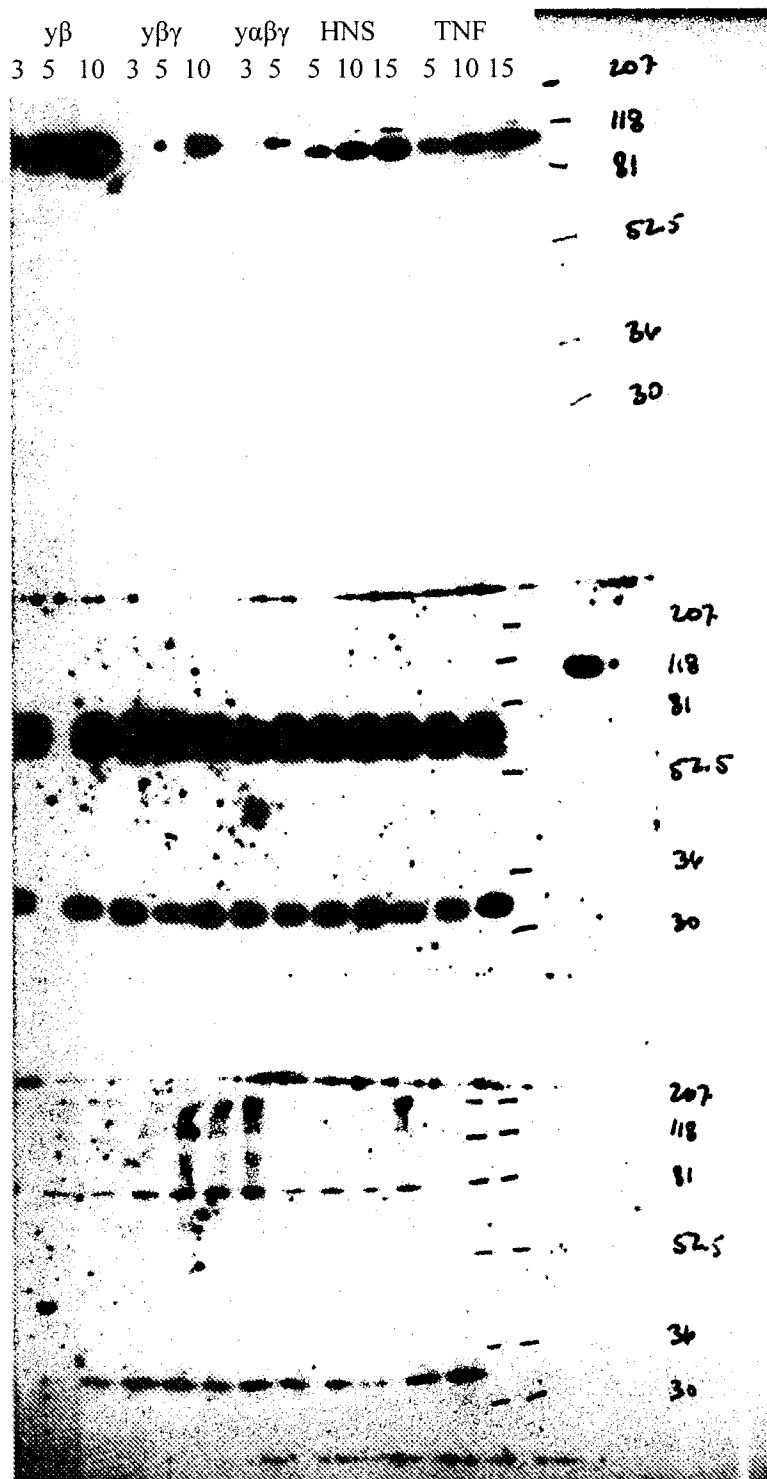
↑

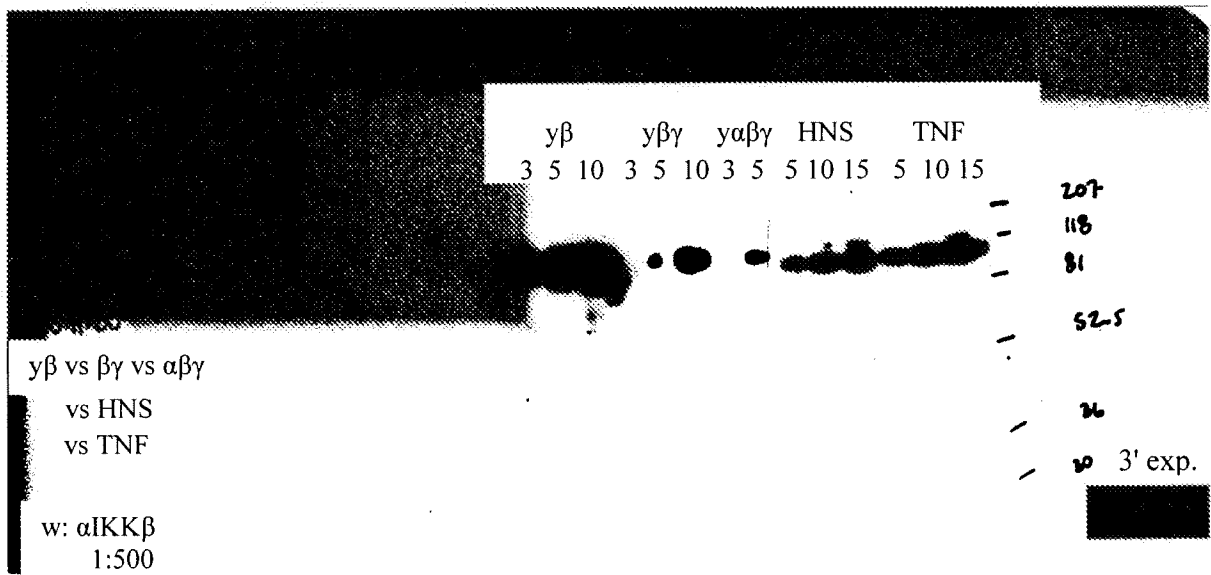
↑ ↑
 range 1-10,000

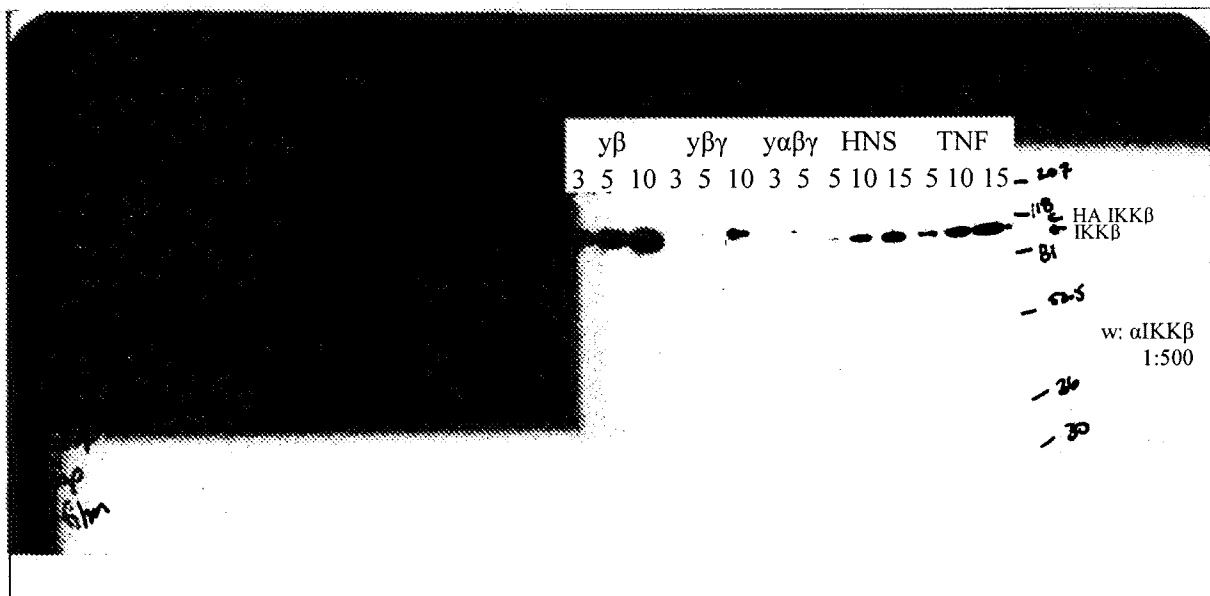
File/Range: D:\Users\1012bsm.gel / 0.000-45853 Counts / 1.000000
User Name: phospho
Image Name: D:\Users\1012bsm.gel
Image Comment: yeast b bg abg HNS TNF-Hela
scanned 9:13 am to 2:05 pm

Present Date/Time:
Scan Date/Time:
Prep. Date/Time:









Purpose: to compare IKK activity in
 $\gamma\beta$ vs $\gamma\beta\gamma$ vs $\gamma\alpha\beta\gamma$ vs HNS vs TNF-Hela

repeat of 10-11 with attempt to use more similar amounts

HNS + TNF (Q20 \rightarrow sup6 GF 10 + 11)

Put 300 λ 1x 1 kinase buffer in bottom to prevent drying.

Top: 200 λ sup6 GF 10 + 200 λ sup6 GF 11

recover ~ 40 λ + adjust vol. to 40 λ
 (1x KA)

β - HA fraction 15

5 λ + 45 λ 1x

Dilute 1:10 in 1x kinase

Tube/lane

2	0.5 λ	5 λ + 16 λ	1x
3	1 λ	10 λ + 11 λ	1x
4	2 λ	20 λ + 1 λ	1x
		equiv. amount	
5	$\beta\gamma$ - 7 λ	~ 5 λ (Hela TNF)	+14 λ 1x
6	14 λ	~10	+7 λ 1x
7	21 λ	~15	+0
8	$\alpha\beta\gamma$ 7	+ 14 λ	1x
9	14	+ 7 λ	1x
10	21	+ 0	
11	HNS	5 + 16 λ	1x
12		10 + 11 λ	1x
13		15 + 6 λ	1x
14	TNF	5 + 16 λ	1x
15		10 + 11 λ	1x
16		15 + 6 λ	1x
17	MW		

1. Aliquot extract + buffer
2. Add 35 λ kinase cocktail Inc 30' 30° C
3. Add 12.2 λ 6x SDS PAGE Heat 95° C 5'
4. Load 10% gel (40 λ)

Cocktail	16 sample	+ 4
10 \times Kinase	48 λ	12 λ
20mm DTT	48 λ	12 λ
200 μ m ATP	48 λ	12 λ
GST - IK $\beta\alpha$	32 λ	8 λ
³² P ATP	8 λ 906-58	2 λ
H ₂ O	<u>296</u>	<u>74</u>
	480	120

20mm DTT
 .02 ml 1M + .98ml H₂O

21 all loaded correctly! 40 λ each

35

56

File/Range: D:\Users\1017bsm.gel / 0.000-45853 Counts / 0.814331

User Name: phospho

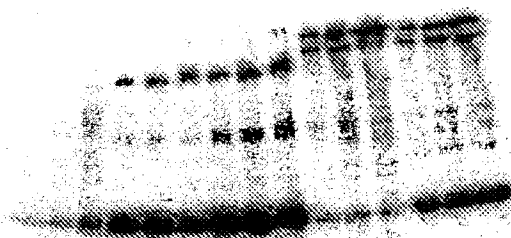
Image Name: D:\Users\1017bsm.gel

Image Comment: 2 experiments
1. 3 M urea GF column fractions (concentrated)
2. yeast b, bg, sbg, HNS, TNF stim Hela

Present Date/Time:

Scan Date/Time:

Prep. Date/Time:



scale 1-250

